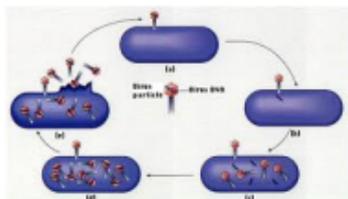


**Directed evolution of phage lysins:
using mathematical models to explore
feasibility/design of new antibacterial drugs**

Jim Bull, Cameron Crandall, Anna Rodriguez, Steve Krone

- Antibiotic resistance \implies search for novel methods to combat bacterial infections
- Directed evolution

Phage lysins



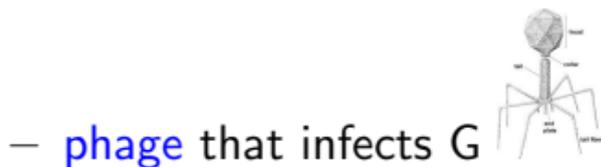
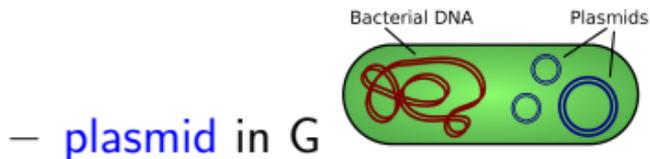
- Lysins have evolved to kill host cell from inside and not harm future hosts
- potential for using lysins to “kill from the outside” (Fischetti, Nelson)
- promising as antimicrobials: only harm bacterial cells; specific bacterial targets; no bacterial resistance
- obstacles to therapeutic use: short half life, etc.

Evolving a better lysin

- Mathematical models: explore feasibility of experimentally evolving better lysins (longer half life, toxicity, diffusivity, adsorption, ...)
- Goal: Improve killing of pathogen **R** (e.g., group A strep) by lysin/toxin
- *Experimental setup: rigged so that “producer” strain **G** benefits from enhanced killing of competitor **R**

Two possible delivery systems

- toxin (that kills R) carried on gene in

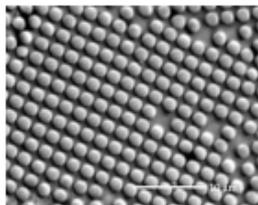


Experimental system

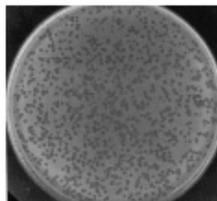
- co-culture R + WT G and/or mutant strain(s) G' (standing genetic variation)
 - select or screen for G'
 - 10 incubation cycles involving R + (G and/or G')
 - **Selection:** after each incubation, collect G-types (or phage) together and resample to start next round of competitions with R
 - **Screening:** only keep samples with “enough G” (fluorescence for sorting)

**** simulation

- Some type of **spatial structure** needed to localize the benefit; in liquid, G and G' would both benefit from improved toxin in G'
 - **emulsion droplets** (≤ 100 cells, G+R or G'+R) for mini competitions



- **agar plates**

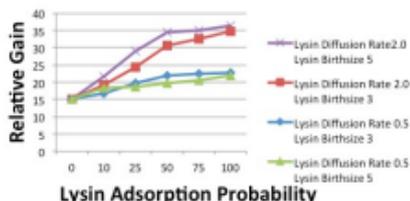
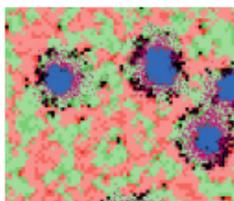


Mathematical Models

Emulsion droplets (endpoint analysis, ODEs): mean q Poisson distribution for initializing G-types in droplets \implies ratio of G' to G after one incubation cycle is

$$\frac{G'_1}{G_1} = \frac{h'}{h} \cdot \frac{G'_0}{G_0} \cdot \frac{2 + q}{2 + q + qG'_0 \left(\frac{h'}{h} - 1 \right)}$$

Plate growth (spatial agent-based models):



Emulsion droplets: Endpoint analysis

Ignore details of what goes on in droplets. Focus on outcome of each incubation:

- Droplets with only G (+R) produce h G during incubation
- Droplets with only G' (+R) produce h' G' during incubation
- Assume (for now) ... only these “pure” droplets

Let p_0 = initial frequency of G' and p_k = frequency at the end of cycle k of G' among all ‘G-types’ ($G' + G$) in the population. Evolutionary dynamics has a simple form:

$$\frac{p_1}{1 - p_1} = \frac{h'}{h} \left(\frac{p_0}{1 - p_0} \right) \quad (1)$$

for the first cycle, and

$$\frac{p_k}{1 - p_k} = \left(\frac{h'}{h} \right)^k \left(\frac{p_0}{1 - p_0} \right) \quad (2)$$

after k cycles. Relevant measure of mutant fitness (W) in this protocol is simply

$$W = \frac{h'}{h}. \quad (3)$$

accounting for mixed droplets

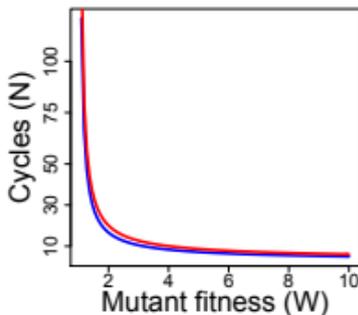
Mean q Poisson distribution for initializing G-types in droplets \implies ratio of G' to G after one incubation cycle is

$$\frac{p_1}{1 - p_1} = \frac{h'}{h} \cdot \frac{p_0}{1 - p_0} \cdot \frac{2 + q}{2 + q + qp_0\left(\frac{h'}{h} - 1\right)}$$

Not a problem when p_0 small. Previous result is a good approximation.

How many incubation cycles?

$$p_N = \frac{W^N p_0}{W^N p_0 + 1 - p_0}$$
$$N [\ln(W)] = \ln \left[\frac{p_N(1 - p_0)}{p_0(1 - p_N)} \right]$$



To go from initial frequency of 10^{-7} to detectable levels (0.01 blue; 0.1 red)

Dynamics within droplet for plasmid model

$$\dot{G} = \psi_G G \left(1 - \frac{G+R}{K}\right)$$

$$\dot{R} = \psi_R R \left(1 - \frac{G+R}{K}\right) - \gamma RL$$

$$\dot{L} = \alpha G \left(1 - \frac{G+R}{K}\right) - \delta L$$

G outgrows R if

$$L > \frac{(\psi_R - \psi_G) \left(1 - \frac{G+R}{K}\right)}{\gamma},$$

but both may be increasing. R decreases if

$$L > \frac{\psi_R \left(1 - \frac{G+R}{K}\right)}{\gamma}.$$

Dynamics within droplet for phage model

$$\dot{P} = -a_1PG + ba_1P_\tau G_\tau$$

$$\dot{G} = -a_1PG + \psi_G G \left(1 - \frac{G + R}{K}\right)$$

$$\dot{R} = \psi_R R \left(1 - \frac{G + R}{K}\right) - a_2LR$$

$$\dot{L} = Za_1P_\tau G_\tau - a_2LR - \delta L$$

Dynamics within droplet for phage model

$$\dot{P} = -a_1 PG + ba_1 P_\tau G_\tau$$

$$\dot{G} = -a_1 PG + \psi_G G \left(1 - \frac{G + R}{K}\right)$$

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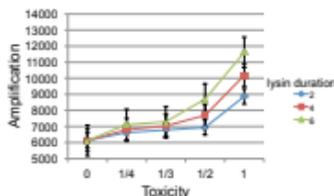
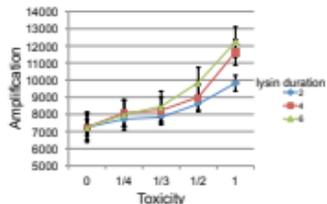
FAILURE: Phage reproduction much faster than bacterial growth. G can't take advantage of removed R.

For phage-delivered lysin to work, need real spatial structure (plates).

Individual-based models of plate growth

Phage model

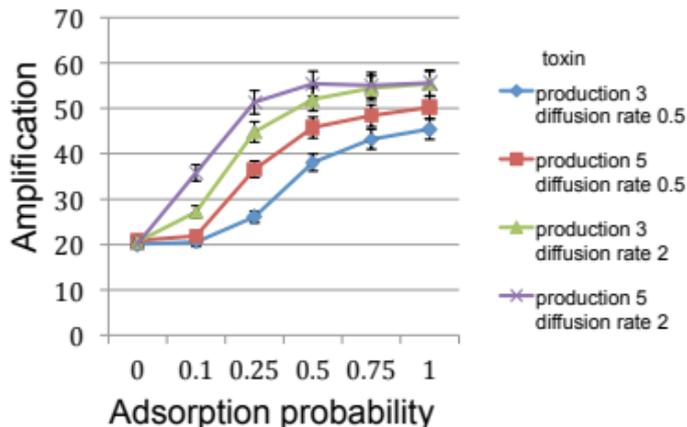
***simulation



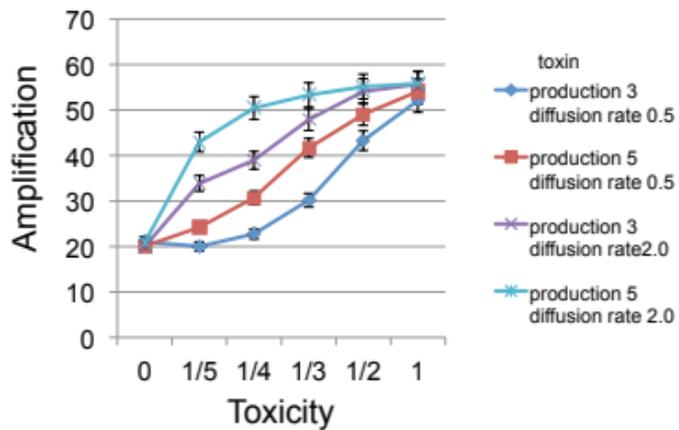
Amplification in number of phage as a function of **lysin toxicity** and **duration** for fast (top) and slow (bottom) phage reproduction; 1.7-1.9X increase. [0 point \implies no lysin]

Individual-based models of plate growth

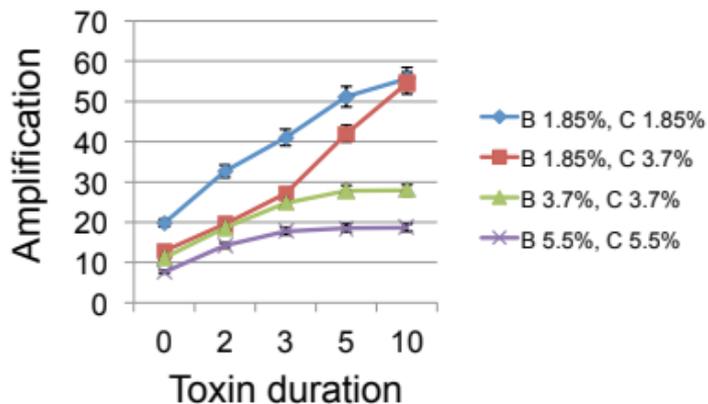
Plasmid model



Amplification in number of G. Select for **faster toxin diffusion**, enhanced production, **higher adsorption rate**.
Overall amplification 2.8X.



Amplification in number of G as function of **toxicity**. Overall amplification 2.7X.



Amplification in number of G as function of **toxin half life**. Larger dynamic range with lower initial cell densities and more R than G. Overall amplification 2.8-4.3X.

Bull et al. (2015), Models for the directed evolution of bacterial allelopathy: bacteriophage lysins. PeerJ 3:e879; DOI 10.7717/peerj.879

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